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**Research Article** 

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# Evaluation of dynamic thiol-disulfide homeostasis on HPV positive-women in progression to cervical intraepithelial lesion

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#### Abstract

Dynamic thiol disulfide homeostasis (TDH) is critical in cervical carcinogenesis at HPV infection as a sign of antioxidant consumption native and total thiol levels decrease in progress to cervical intraepithelial lesions. TDH is the main actor in signaling pathways, apoptosis, antioxidant and detoxification reactions. In this study, we aimed to evaluate the effect of TDH intraepithelial progression of cervical precancerous lesions on HPV positive women. This was a prospective cross-sectional study. Subjects were selected from newly diagnosed high risk HPV DNA-positive patients. TDH results were calculated as the levels of disulfide, native and total thiol, the ratios of disulfide/total thiol (SS/SH+SS), disulfide/native thiol (SS/SH) and native thiol/total thiol (SH/SH+SS). A total of 146 women were included in the study. Study groups were as group one; control included 66 participants, group two; HPV DNA-positive women without preinvasive cervical lesion included 50 participants. Native and total thiol levels were elevated on HPV-positive women without preinvasive cervical lesions. There were no significant differences between groups related to the ratios of SS/SH, SS/ Total SH, SH/ Total SH levels. HPV infection related to oxidative stress has effects on oxidant/antioxidant balance and could be demonstrated in systemic circulation by TDH parameters. Consumption of thiol substances play a role in the cervical neoplastic process, replacement with antioxidants would be a treatment option for HPV infections.

Keywords: antioxidants, cervical intraepithelial lesions, human papillomavirus, oxidative stress, thiol-disulfide homeostasis

## 1. Introduction

Human Papilloma Virus (HPV) is a double-stranded DNA virus that involves the squamous epithelium. HPV is a sexual transmitted infection and the main cause of the cervical cancer. There are about 40 HPV types indicating anogenital involvement. High oncogenic types of HPV persistence cause cervical intraepithelial lesions and cancer (CIN) formation. More than 90% of HPV infection is cleared from the body in about two years (1-3). On the other hand, up to 10% of the infection persists and causes a cervical intraepithelial lesion.

HPV targets basal layer and metaplastic cells to create infection and reaches there through the micro abrasions formed in the stratified squamous epithelium. After transmission of the virus to the cervical epithelium, it integrates host genome and oncogenic differentiation occurs by expressing E6 and E7 oncogenes in the cell. Dysfunction at native immune response of the host, chronic inflammation and oxidative stress also have an effect on HPV persistence and carcinogenesis. Impairment at oxidant-antioxidant balance and elevated oxidative status were found to be correlated with CIN and cervical carcinoma (4, 5).

The primary target of oxygen radicals are proteins such as cysteine, methionine, glutathione called sulfides containing sulfide groups. These proteins oxidize to form reversible disulfide bonds. Structural and functional changes occur in these proteins during losing thiol groups (6, 7). Plasma and tissue levels of thiol groups decrease in the course of prevention from the destructive effects of free oxygen radicals (8). In many cellular events as signaling pathways, apoptosis, antioxidant and detoxification reactions, dynamic TDH is the main actor (9, 10).

In recent years, TDH has maintained its popularity and has been the subject of many studies. In the literature, there are a growing body of studies showing the effect of TDH in many acute and chronic disorders. In this study, we evaluated the effect of TDH intraepithelial progression of cervical precancerous lesions at HPV-positive women. This is the first study that investigates the potential impact of TDH on cervical carcinogenesis.

# 2. Material and Method

This prospective cross-sectional study was performed in the Gynecology and Obstetrics Department of Trabzon Kanuni Training and Research Hospital. The study was organized in accordance with the Helsinki Declaration guide and received approval from the local ethics committee of Trabzon Kanuni Training and Research Hospital (23618724-799, 2022/01). The informed consent form was subscribed by all participants after giving information about the study.

A total of 146 women were included in the study. The participants consist of three groups. Group one was control group including 66 participants (with negative HPV and pap smear test), group two was HPV DNA positive women without preinvasive cervical lesion and included 30 participants and group three was HPV DNA positive-women with preinvasive cervical lesion and included 50 participants. Age, gravidity, body mass index (BMI) were evaluated as demographic features.

Subjects were selected from newly diagnosed high-risk HPV DNA-positive patients without concomitant active sexually transmitted infections, previous history of cervical preinvasive lesions or cervical cancer. Pregnancy, lactation, smoking, any disease associated with immune deficiency, corticosteroid usage, having any genitourinary system infection were the other exclusion criteria. Age matched women who were admitted for routine gynecological checkups were enrolled as a control group. All participants were interrogated about sexual, reproductive, medical and surgical history.

Gynecological evaluation was performed including bimanual palpation and transvaginal ultrasonographic assessment. During speculum examination cervicovaginal swaps obtained for Liquid-based (ThinPrep Pap Test, Hologic) cervicovaginal smear test and the Hybrid Capture 2 DNA test (Qiagen, Hilden, Germany) for High-Risk HPV detection. Bethesda 2001 classification system was used in the evaluation of smear tests. HPV carriers underwent to colposcopic evaluation (Leica MSV 197, Germany), 3-5% acetic acid administered for visualization of cervical intraepithelial lesions and punch biopsies were taken from suspected areas.

About 4 mL of fasting blood picked up by venopuncture from the antecubital region. The blood samples were centrifuged at 4000 rpm for 10 minutes, and the serum was decomposed and stored at  $-80^{\circ}$ C until an assessment of TDH. Serum TDH was evaluated with an automated spectrophotometric measuring technique defined by Erel (11). TDH results were figured out as  $\mu$ mol/L.

Firstly, disulfide links were reduced by using sodium borohydride to form free functional thiol groups. For prevention of the reduction of 5,5'-dithiobis-(2-nitrobenzoic)

acid (DTNB) reductant sodium borohydride was consumed and removed with formaldehyde. Reduced and native thiol groups were identified after the reaction with DTNB. The dynamic disulfide amount provided from the difference between the total and native thiols and calculated by the relationship between disulfide and thiol groups. If the ratio increases in favor of thiols, it indicates oxidative stress, while an increase in favor of disulfides indicates elevation of antioxidant capacity. The parameters for calculation of dynamic TDH were disulfide/total thiol percent ratios (SS/SH+SS), disulfide/native thiol percent ratios (SS/SH), and native thiol/total thiol percent ratios (SH/SH+SS), disulfide, native and total thiol quantity.

## 2.1. Statistical analysis

Version 18 of SPSS was used for statistical analysis. Kolmogorov-Smirnov method was used to determine the normal distribution of data. Mean  $\pm$  standard deviation was used for normally distributed data, and median  $\pm$  interquartile range values were used for non-parametric data. Comparison of TDH between study groups was done with One Way ANOVA test with post hoc LSD analysis.

# 3. Results

Mean age of the participants was  $42.22\pm7.16$  year, gravidity  $3.6\pm0.61$  and BMI  $25.56\pm3.27$  kg/m<sup>2</sup>. No significant differences were obtained amongst the groups (p>0.05).

TDH was measured by native thiol and disulfide values and SS/ total SH, SH/ total SH, SS/ SH ratios. Mean native thiol levels were 246.84±81.80 (µmol/L), disulfide 22.07±7.26 (µmol/L), SS/ total SH ratio was 8.21±3.81 (µmol/L), SH/ total SH ratio was 83.52±7.63 (µmol/L) and SS/ SH ratio was 10.49±7.67 (µmol/L). Comparison of TDH parameters between study groups was given in Table 1. There were no significant differences between group according to native thiol, disulfide, SS/SH, SS/ Total SH, SH/ Total SH levels. Native and total thiol levels were elevated in HPV positive women without preinvasive cervical lesions. Contrarily in cervical intraepithelial lesion group native and total thiol levels were decreased, however the results could not reach to statistical significance (Fig. 1).

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I able 1.	I hiol	disulfide	homeostasis	parameters	ın	study	groups

	Group 1 (n=66)	Group 2 (n=30)	Group 3 (n=50)	Р
Native Thiol (µmol/L)	249,96±74,06	273,26±68,45	233,21±82,72	0,07
Total Thiol (μmol/L)	293,96±80,14	317,86±68,17	271,08±93,52	0,05
Disulfide (µmol/L)	22,84±7,68	22,29±5,76	21,31±6,85	0,51
SS/SH (µmol/L)	10,05±5	8,90±3,69	10,75±5,76	0,29
SS/Total SH (µmol/L)	8,14±3,20	7,40±2,58	8,70±3,98	0,25
SH/Total SH (µmol/L)	83,81±6,52	85,18±5,17	82,35±8,67	0,21

Data are given mean±std; (-SH): sulfhydryl



Fig. 1. The graph of total thiol levels distribution in study groups

#### 4. Discussion

In this study, we compared the differences between TDH parameters, which are the main components of antioxidant protection, in HPV infected women without cervical intraepithelial lesions and those who developed cervical dysplasia.

Our results demonstrated that TDH parameters affected in HPV positive- women with cervical intraepithelial lesions. Evaluations from peripheral blood samples of HPV positivewomen represented that total and native thiol levels decreased at cervical intraepithelial lesions. Our findings supported that HPV infection related to cellular changes activates detoxification systems and causes consumption of cellular antioxidant. However, our results were not statistically significant, differences between study groups were thought that cervical neoplastic progression effects oxidant/antioxidant status and that could be shown in systemic circulation. These findings demonstrated that HPV infection caused cervical neoplastic process is closely related with oxidant and antioxidant regulation. According to this data we concluded that replacement of antioxidants in HPV infection could be a strategy for treatment of infection and prevention from cellular changes.

Experimental and observational studies show that a large portion of HPV infection are spontaneously regressed however a small part of them progress and generate cervical cancer. Being HPV infected is not just enough for cancer development, there is still lack of information about other individual and environmental factors and their mechanisms on pathogenesis. Free oxygen radicals are important factors, those are effective in carcinogenesis by signaling pathway up regulation, cell differentiation, proliferation and change of cellular survival. HPV reproduces in infected and transformed cells and disrupts the redox balance (12-15). Past studies showed that immune response of the host to HPV infection influences oxidative stress and could be demonstrated with alteration of stress markers. Siegel et al (16) evaluated the relation between oxidant load and HPV clearance, and they determined a high oxidant status. On the contrary, an increase in oxidant levels, antioxidant enzymes have been shown to be lowered in patients with CIN and cervical carcinoma due to excessive consumption (17-19). According to an in-vivo study evaluating the effect of the Redox system on carcinogenesis, an increased oxidant environment has been shown to be effective in HPV 16 neoplastic progression, and oxidative modification of DNA and proteins in dysplastic tissues have influenced cellular differentiation, leading to neoplastic progression. In cancerous tissue, controlling oxidative damage could be provided by selective reduction of key detoxification proteins (13).

Histopathological evaluations show an increased inflammatory infiltration in severe HPV-induced lesions. In the early stages, the infection caused by the virus at basal cells is not associated with circulating immune cells therefore inflammation does not play a central role in the pathogenesis of HPV infection. Persistent infection causes chronic inflammation and triggers an imbalance between pro-oxidant and antioxidants (14).

During intracellular reactive oxygen species increase, local antioxidant capacity and numerous intracellular adaptive mechanisms upregulate to prevent the development of apoptosis and to protect the tissue. Throughout free radicals rise above physiological levels, the regulation of redox homeostasis, which was the cellular protection system of the organism, is disrupted and initiates the process of uncontrolled cell growth and carcinogenesis (20, 21). Redox homeostasis is controlled by oxidizing and reducing of free radicals and thiolcontaining proteins in the cell.

Thiols are the parts of the natural antioxidant enzyme system in the organism that contain Sulfhydryl and form disulfide in antioxidant activation (22). Attachment of sulfur and hydrogen atoms to a carbon atom forms sulfhydryl and oxidation reactions form disulfide bonds between two sulfhydryl groups (23). This binding is reversible and disulfides can reduce to thiol groups to sustain homeostasis (24). This homeostasis plays a crucial role in antioxidant protection, detoxification, signal transmission, programmed cell death, organizing enzymatic reaction, transcription factors and intra and inter-cellular signaling mechanisms (25).

Many compounds like cysteine, methionine, glutathione, homocysteine, cysteinyl-glycine and glutamyl cysteine are containing thiol groups and have structural alterations under oxidative stress. These proteins oxidize to form reversible disulfide bonds. Structural and functional changes occur in these proteins during losing thiol groups (6, 7). Plasma and tissue levels of thiol groups decrease in the course of prevention from the destructive effects of free oxygen radicals (8). The transformation of thiols into disulfides is an early indicator of protein oxidation from reactive oxygen radicals. Measurement of total thiol level and determination of TDH are the mirror of excessive free oxygen radical formation in many

## illnesses (26).

Disruption of this ratio acts a part in the pathogenesis of many inflammatory diseases such as diabetes mellitus, inflammatory arthritis, renal failure, cancer, Parkinson, Alzheimer, multiple sclerosis. Shifting thiol/disulfide balance to disulfide direction was seen in degenerative diseases such as diabetes, obesity, and pneumonia, to thiol direction poses risk factor neoplastic processes such as multiple myeloma, bladder, colon and kidney cancer (27).

The colorimetric method improved by Erel and Neşelioğlu (11) renders possible to provide information about oxidative stress by identifying the total plasma thiol/disulfide ratio. The easy, inexpensive and practical method is carried out with a fully automatic analyzer that does not require separation. It can be used to evaluate free radicals synthesized by many different metabolic pathways, including aerobic respiration in mitochondria (12). Before this measurement technique was developed, only low molecular weight thiol components which were cysteine, glutathione, and homocysteine could be measured. This method allows measuring the majority part of thiol and disulfides in albumin and proteins. According to the old method, thiol / disulfide measurement did not reflect true homeostasis.

In enzymes containing thiol, free radicals formed after normal metabolites or pathological processes cause structural and functional disturbance and alterations in thiol/disulfide balance. A decrease in plasma thiol concentration indicates an increase in free radical formation. A small proportion of HPVinfected cells progresses to cancer, the expression of E6 and E7 oncogenes play roles in this process. Camporeale et al. (28) studied the molecular mechanism of potential damages of the oxidative environment in HPV infected cells. They reported that carboxy-terminal of E7 oncoprotein is rich in the domain of cysteine and sensitive to ROS. Exposure to free radicals regulates the transition from the cytoplasm to the nucleus by creating disulfide bonds.

HPV infection has a local effect at cervical epithelium and evaluation of TDH parameters at cervical secretions could be most informative for detection of oxidant/ antioxidant status. This was one of the limitations of our study. The other limitation was cross-sectional design of the study, the levels of TDH parameters previous the infection and cervical lesion were not included in the study.

In conclusion, HPV infection related oxidative stress has systemic effects and could be demonstrated in the systemic circulation by TDH parameters. Consumption of thiol substances play a role in cervical neoplastic process, replacement with antioxidants would be a treatment option for HPV infections.

## **Conflict of interest**

The authors declared no conflict of interest.

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# Authors' contributions

Concept: Y.B.T., Design: Y.B.T., Data Collection or Processing: Y.B.T., R.E., H.T., Analysis or Interpretation: Y.B.T., Ö.E., Literature Search: Y.B.T., R.E., Writing: Y.B.T, R.E.

## References

- Myers ER, McCrory DC, Nanda K, Bastian L, Matchar DB. Mathematical model for the natural history of human papillomavirus infection and cervical carcinogenesis. Am J Epidemiol. 2000;15;151(12):1158-71. doi: 10.1093/oxfordjournals.aje.a010166.
- 2. Richardson H, Kelsall G, Tellier P, Voyer H, Abrahamowicz M, Ferenczy A et al. The natural history of type-specific human papillomavirus infections in female university students. Cancer Epidemiol Biomarkers Prev. 2003;12(6):485-90. PMID: 12814991.
- 3. Plummer M, Schiffman M, Castle PE, Maucort-Boulch D, Wheeler CM; ALTS Group. A 2-year prospective study of human papillomavirus persistence among women with a cytological diagnosis of atypical squamous cells of undetermined significance or low-grade squamous intraepithelial lesion. J Infect Dis. 2007;1;195(11):1582-9. doi: 10.1086/516784.
- Looi ML, Mohd Dali AZ, Md Ali SA, Wan Ngah WZ, Mohd Yusof YA. Oxidative damage and antioxidant status in patients with cervical intraepithelial neoplasia and carcinoma of the cervix. Eur J Cancer Prev. 2008;17(6):555-60. doi: 10.1097/CEJ.0b013e328305a10b.
- Kim SY, Kim JW, Ko YS, Koo JE, Chung HY, Lee-Kim YC. Changes in lipid peroxidation and antioxidant trace elements in serum of women with cervical intraepithelial neoplasia and invasive cancer. Nutr Cancer. 2003;47(2):126-30. doi: 10.1207/s15327914nc4702 3.
- 6. Dean RT, Fu S, Stocker R, Davies MJ. Biochemistry and pathology of radical-mediated protein oxidation. Biochem J. 1997;15;324 (Pt 1)(Pt 1):1-18. doi: 10.1042/bj3240001.
- 7. Ziegler DM. Role of reversible oxidation-reduction of enzyme thiols-disulfides in metabolic regulation. Annu Rev Biochem. 1995;54:305-29. doi: 10.1146/annurev.bi.54.070185.001513.
- McCord JM. Human disease, free radicals, and the oxidant/antioxidant balance. Clin Biochem. 1993; 26(5):351-7. doi: 10.1016/0009-9120(93)90111-i.
- **9.** Biswas S, Chida AS, Rahman I. Redox modifications of proteinthiols: emerging roles in cell signaling. Biochem Pharmacol. 2006;28;71(5):551-64. doi: 10.1016/j.bcp.2005.10.044.
- **10.** Circu ML, Aw TY. Reactive oxygen species, cellular redox systems, and apoptosis. Free Radic Biol Med. 2010;15;48(6):749-62. doi: 10.1016/j.freeradbiomed.2009.12.022.
- 11. Erel O, Neselioglu S. A novel and automated assay for thiol/disulphide homeostasis. Clin Biochem. 2014;47(18):326-32. doi: 10.1016/j.clinbiochem.2014.09.026.
- **12.** De Marco F. Oxidative stress and HPV carcinogenesis. Viruses. 2013;12;5(2):708-31. doi: 10.3390/v5020708.
- 13. De Marco F, Bucaj E, Foppoli C, Fiorini A, Blarzino C, Filipi K,

et al. Oxidative stress in HPV-driven viral carcinogenesis: redox proteomics analysis of HPV-16 dysplastic and neoplastic tissues. PLoS One. 2012;7(3):e34366. doi: 10.1371/journal.pone.0034366.

- 14. Williams VM, Filippova M, Soto U, Duerksen-Hughes PJ. HPV-DNA integration and carcinogenesis: putative roles for inflammation and oxidative stress. Future Virol. 2011;1;6(1):45-57. doi: 10.2217/fvl.10.73.
- Foppoli C, De Marco F, Cini C, Perluigi M. Redox control of viral carcinogenesis: The human papillomavirus paradigm. Biochim Biophys Acta. 2015;1850(8):1622-32. doi: 10.1016/j.bbagen.2014.12.016.
- 16. Siegel EM, Patel N, Lu B, Lee JH, Nyitray AG, Craft NE, et al. Biomarkers of oxidant load and type-specific clearance of prevalent oncogenic human papillomavirus infection: markers of immune response? Int J Cancer. 2012;1;131(1):219-28. doi: 10.1002/ijc.26363.
- 17. Sedjo RL, Roe DJ, Abrahamsen M, Harris RB, Craft N, Baldwin S, et al. Vitamin A, carotenoids, and risk of persistent oncogenic human papillomavirus infection. Cancer Epidemiol Biomarkers. 2002;11(9):876-84. PMID: 12223432.
- 18. Manju V, Kalaivani Sailaja J, Nalini N. Circulating lipid peroxidation and antioxidant status in cervical cancer patients: a case-control study. Clin Biochem. 2002;35(8):621-5. doi: 10.1016/s0009-9120(02)00376-4.
- 19. Looi ML, Mohd Dali AZ, Ali SA, Wan Ngah WZ, Mohd Yusof YA. Oxidative damage and antioxidant status in patients with cervical intraepithelial neoplasia and carcinoma of the cervix. Eur J Cancer Prev. 2008;17(6):555-60. doi: 10.1097/CEJ.0b013e328305a10b.
- 20. Burhans WC, Weinberger M. DNA replication stress, genome

instability and aging. Nucleic Acids Res. 2007;35(22):7545-56. doi: 10.1093/nar/gkm1059.

- 21. Khansari N, Shakiba Y, Mahmoudi M. Chronic inflammation and oxidative stress as a major cause of age-related diseases and cancer. Recent Pat Inflamm Allergy Drug Discov. 2009;3(1):73-80. doi: 10.2174/187221309787158371.
- 22. Cremers CM, Jakob U. Oxidant sensing by reversible disulfide bond formation. J Biol Chem. 2013;13;288(37):26489-96. doi: 10.1074/jbc.R113.462929.
- **23.** Jones DP, Liang Y. Measuring the poise of thiol/disulfide couples in vivo. Free Radic Biol Med. 2009;15;47(10):1329-38. doi: 10.1016/j.freeradbiomed.2009.08.021.
- 24. Biswas S, Chida AS, Rahman I. Redox modifications of proteinthiols: emerging roles in cell signaling. Biochem Pharmacol. 2006;28;71(5):551-64. doi: 10.1016/j.bcp.2005.10.044.
- **25.** Circu ML, Aw TY. Reactive oxygen species, cellular redox systems, and apoptosis. Free Radic Biol Med. 2010;15;48(6):749-62. doi: 10.1016/j.freeradbiomed.2009.12.022.
- **26.** Ozyazici S, Karateke F, Turan U, Kuvvetli A, Kilavuz H, Karakaya B, et al. A Novel Oxidative Stress Mediator in Acute Appendicitis: Thiol/Disulphide Homeostasis. Mediators Inflamm. 2016;2016:6761050. doi: 10.1155/2016/6761050.
- 27. Aksoy M, Çelik H. Dynamic thiol/disulphide homeostasis in vitiligo patients. Postepy Dermatol Alergol. 2018;35(5):498-501. doi: 10.5114/ada.2018.72856.
- 28. Camporeale G, Lorenzo JR, Thomas MG, Salvatierra E, Borkosky SS, Risso MG, et al. Degenerate cysteine patterns mediate two redox sensing mechanisms in the papillomavirus E7 oncoprotein. Redox Biol. 2017;11:38-50. doi: 10.1016/j.redox.2016.10.020.